**Oligodendrocyte Staining**

**Day 1**

1. PFA 30’
2. PBS 3x5’
   1. Now option to leave in fridge in PBS
3. Transfer to new plates
4. NH4Cl 60’
   1. 3x20’ if the plate is old, to wash away nead cells
5. Saturate in serum buffer 30-120’
   1. Serum buffer: 10% DS, 3% BSA, 0.01% PBS-T (for 50mL: 5mL DS, 1.5g BSA, 1.25mL Triton-X100 with cut tip)
6. Make antibody master mix in serum buffer and then add 0.5mL to each well
7. Cold room O/N

**Day Two**

1. PBS 3x15’
2. Make master mix of secondaries
3. Shake for a couple hours, away from light
4. PBS 3x15’
5. If for STORM:
   1. Cold 4% PFA 5-10’
   2. PBS 3x30’
6. If for confocal:
   1. Hoechst 30-40’, then return to bottle, dark
   2. PBS 2x7’
   3. Mount

Tubulin (mouse) 1:1000

Bin1 BSH3 (rabbit) 1:1000

GFP (chicken) 1:5000

MBP (rat) 1:1000

CNP (mouse) 1:500

WASP (rabbit) 1:500